# (-)-N-[<sup>11</sup>C]Propyl-norapomorphine [<sup>11</sup>C]NPA

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Chemical name: (-)-N-[11C]Propyl-

norapomorphine

Abbreviated name: [11C]NPA

Synonym:

Backbone: Compound

**Target:** D<sub>2</sub> dopamine receptors

Mechanism: Receptor binding

Method of detection: PET
Source of signal: 11C
Activation: No
In vitro studies: Yes
Rodent studies: Yes
Other non-primate manual

studies:

Human studies: No

Non-human primate studies: Yes

H-C-H

Click on the above structure for additional information in PubChem [http://pubchem.ncbi.nlm.nih.gov].

## **Background**

#### [PubMed]

Dopamine, a neurotransmitter, plays an important role in the mediation of movement, cognition, and emotion (1, 2). Dopamine receptors are involved in the pathophysiology of neuropsychiatric diseases, such as Parkinson's disease, Alzheimer's disease, Huntington's disease, and schizophrenia (3). Five subtypes of dopamine receptors,  $D_1$  through  $D_5$ , have been well characterized pharmacologically and biochemically (4). These five subtypes are classified into two subfamilies:  $D_1$ -like ( $D_1$  and  $D_5$ ) and  $D_2$ -like ( $D_2$ ,  $D_3$ , and  $D_4$ ) dopamine receptors.  $D_1$ -like and  $D_2$ -like receptors exert synergistic as well as opposite effects at both the biochemical and overall system level. A great majority of striatal  $D_1$  and  $D_2$  receptors are localized postsynaptically on caudate-putamen neurons and to a lesser extent presynaptically on nigrostriatal axons.

Dopamine receptors are G-protein-coupled receptors and exist in high- and low-affinity states, with respect to agonist binding. The two states are interconvertible. In the high-affinity state, dopamine receptors are coupled to G-proteins, whereas the low-affinity state they are not. Dopamine has a dissociation constant ( $K_d$ ) of 7 nM for the high-affinity state ( $K_{high}$ ) and a  $K_d$  of 1,720 nM for the low-affinity state ( $K_{low}$ ) (5). Under physiologic conditions, dopamine is expected to bind predominately to receptors in the high-affinity state. The high-affinity state was suggested to be the functional form of the dopamine receptors.

Substituted benzamides, such as sulpiride, raclopride, and iodobenzamide, are specific ligands with only moderate affinity for the  $D_2$  receptors, making studies of extrastriatal  $D_2$  receptors difficult (6-8). In binding studies, <sup>123</sup>I-labeled epidepride, an analog of isoremoxipride, was found to have high potency and low nonspecific binding and to be selective for striatal and extrastriatal  $D_2$  receptors (9). Epidepride has marginal binding to  $D_4$  receptors, with little affinity for other known neurotransmitter receptors. (S)-*N*-((1-AllyI-2-pyrrolidinyI)methyI)-5-(3-[<sup>18</sup>F]fluoropropyI)-2,3-dimethoxybenzamide ([<sup>18</sup>F]fallypride [http://www.ncbi.nlm.nih.gov/books/bv.fcgi? rid=micad.chapter.Fallypride18F]), an analog of epidepride, was found to be a selective, high-affinity antagonist of  $D_{2/3}$  receptors (10), and in positron emission tomography (PET) *in vivo* studies (11-13), it identified extrastriatal  $D_{2/3}$  receptors. However, none of these antagonists distinguishes between the high- and low- affinity states of the  $D_2$  receptors. (–)-*N*-Propyl-norapomorphine (NPA) was reported to have  $K_{high}$  and  $K_{low}$  values of 0.07-0.4 and 20-200 nM, respectively (5, 14-16). This provides a >50-fold selectivity for high-affinity over low-affinity receptors. NPA has good affinity ( $K_i = 0.3$  nM) for  $D_3$  receptors but not other neurotransmitters (17). [<sup>11</sup>C]NPA is being developed as a PET agent for the non-invasive study of the high-affinity state of the  $D_{2/3}$  receptors in the brain.

## **Synthesis**

#### [PubMed]

Wong et al. (18) reported a one-pot synthesis of [11C]NPA in which [11C]propionyl chloride was reacted with norapomorphine followed by LiAlH<sub>4</sub> reduction, with a radiochemical yield of 16% (based on [11C]CO<sub>2</sub> at the end of bombardment) and an average specific activity of 63 GBq/µmol (1,700 mCi/µmol at end of synthesis) after purification by C-18 Sep-Pak and high-performance liquid chromatography. Radiochemical purities were >99%. [11C]Propionyl chloride was prepared by reacting [11C]CO<sub>2</sub> with ethylmagnesium bromide, followed by reaction with phthaloyl chloride. The total synthesis time was 60 min.

## In Vitro Studies: Testing in Cells and Tissues

#### [PubMed]

In a binding study of dopamine receptors in membranes of the porcine anterior pituitary, [ $^3$ H] NPA had an average  $K_d$  of 0.26  $\pm$  0.01 nM and a  $B_{max}$  of 2.3  $\pm$  0.1 pmol/g tissue (19). Guanilylimidodiphosphate completely inhibited [ $^3$ H]NPA binding, suggesting that [ $^3$ H]NPA was binding primarily

to dopamine  $D_2$  receptors in the high-affinity state. In the presence of [ $^3$ H]spiroperidol, NPA had  $K_{\text{high}}$  and  $K_{\text{low}}$  values of 0.27 ± 0.04 and 26 ± 2.6 nM, respectively. About 54% of  $D_2$  receptors were in the high-affinity state. Therefore, NPA has good selectivity and affinity for the high-affinity state of  $D_2$  receptors. George et al. (5) reported that NPA had  $K_{\text{high}}$  and  $K_{\text{low}}$  values of 0.31 and 207 nM, respectively.

## **Animal Studies**

#### **Rodents**

#### [PubMed]

Biodistribution studies in rats showed high accumulation of radioactivity in the kidney (1.00% injected dose (ID)/g), followed by the liver (0.72% ID/g), adrenal (0.66% ID/g), lung (0.31% ID/g), and spleen (0.25% ID/g) at 5 min after injection of [11C]NPA (18). There was marked accumulation of the tracer in the striata within the first 30 min (0.88% ID/g), followed by a decrease of radioactivity to 0.56% ID/g at 60 min. The striatum/cerebellum and frontal cortex/cerebellum ratios were 3.47 and 1.44, respectively, at 30 min after injection. Haloperidol pretreatment (1 mg/kg) effectively blocked specific binding of [11C]NPA to the striatum (from 0.88 to 0.28% ID/g) and frontal cortex (from 0.36 to 0.21% ID/g) at 30 min. Little inhibition was seen in the cerebellum (from 0.25 to 0.22% ID/g). [11C]NPA thus displays uptake and wash-out kinetics characteristic of reversible radiotracers.

#### **Other Non-Primate Mammals**

[PubMed]

No publication is currently available.

#### **Non-Human Primates**

#### [PubMed]

[ $^{11}$ C]NPA PET studies in non-human primates have provided useful assessment of the D $_2$  receptor in the brain, showing localization of [ $^{11}$ C]NPA in striatal regions without agonist effects on various physiologic parameters, such as blood pressure, heart rate, respiratory rate, and body temperature (20). Wong et al (18). showed selective uptake in the striatum (0.031% ID/g) of a baboon monkey brain with striatum/cerebellum ratios of 2.33 at 15 min and 2.86 at 45 min after injection of 231 MBq (6.25 mCi) of [ $^{11}$ C]NPA. The striatal accumulation of [ $^{11}$ C]NPA was inhibited by pretreatment with haloperidol (1 mg/kg) with a striatum/cerebellum ratio of 1.29 at 45 min after injection.

Wong et al. (18) also performed quantitative measurements of [ $^{11}$ C]NPA binding with kinetic and graphical analyses, using arterial input function to derive the binding potential (BP) and specific-to-nonspecific equilibrium partition coefficient ( $V_3$ ) in two baboons. In kinetic analyses, BP estimates were 4.04 ± 1.05 ml/g in the striatum, whereas BP estimates were 3.90 ± 1.03 ml/g by graphical analysis with arterial input. At 40 min post injection, 31% of [ $^{11}$ C]NPA radioactivity

remained intact in the arterial plasma. The authors concluded that data from 30 min of scanning were sufficient to derive  $V_3$  values by kinetic, graphical, and simplified reference-tissue model analyses.

Narendran et al. (20) studied 3 male baboons with [11C]raclopride (a D<sub>2</sub> antagonist) and [11C] NPA under baseline conditions and after administration of the potent dopamine releaser amphetamine. Kinetic modeling with an arterial input function was used to derive the striatal V3". The [ $^{11}$ C]raclopride  $V_3$  was reduced by 24 ± 10%, 32 ± 6%, and 44 ± 9% after amphetamine doses of 0.3, 0.5, and 1.0 mg/kg, respectively, whereas the [ $^{11}$ C]NPA  $V_3$ " was reduced by 32 ± 2%, 45 ± 3%, and  $53 \pm 9\%$ , respectively, after the same doses of amphetamine. Thus, endogenous dopamine was 42% more effective at competing with [11C]NPA binding than [11C]raclopride binding, which is consistent with the pharmacology of these tracers (agonist versus antagonist). These results also suggest that 71% of D<sub>2</sub> receptors are configured in a state of high affinity for agonists in vivo. [<sup>11</sup>C] NPA is able to detect the change in dopamine levels induced by D-amphetamine and is more vulnerable to competition by endogenous dopamine than to competition by the antagonist radiotracer [11C]raclopride. Because raclopride binds to receptors in both the high- and low-affinity states and dopamine binds with high affinity mainly to receptors in the high-affinity state, the percentage reduction is greater when an agonist is used than when an antagonist is used. This large proportion of high-affinity sites might explain the vulnerability of D2 radiotracers to competition by endogenous dopamine and is consistent with the reported in vivo binding of the agonist radiotracer [11C]NPA.

Narendran et al. (21) performed further PET studies in 3 baboons under non-carrier- and carrier-added conditions to compare the  $B_{\text{max}}$  values of [ $^{11}$ C]NPA and [ $^{11}$ C]raclopride in the same baboons. The [ $^{11}$ C]raclopride  $K_{\text{d}}$  and  $B_{\text{max}}$  were 1.59 ± 0.28 nM and 27.3 ± 3.9 nM, respectively. The *in vivoK*<sub>d</sub> of [ $^{11}$ C]NPA was 0.16 ± 0.01 nM, consistent with its reported affinity for D<sub>2high</sub>*in vitro* binding (20). The  $B_{\text{max}}$  for [ $^{11}$ C]NPA was 21.6 ± 2.8 nM and 79% of the [ $^{11}$ C]raclopride  $B_{\text{max}}$ . This result suggested that 79% of D<sub>2</sub> receptors are configured in the high-affinity state *in vivo*.

# **Human Studies**

[PubMed]

No relevant publication is currently available.

## References

- Carbon M, Ghilardi MF, Feigin A, Fukuda M, Silvestri G, Mentis MJ, Ghez C, Moeller JR, Eidelberg D. Learning networks in health and Parkinson's disease: reproducibility and treatment effects. Hum Brain Mapp 19(3):197–211; 2003. (PubMed)
- Chesselet MF, Delfs JM. Basal ganglia and movement disorders: an update. Trends Neurosci 19(10):417–422; 1996. (PubMed)
- Seeman P, Bzowej NH, Guan HC, Bergeron C, Reynolds GP, Bird ED, Riederer P, Jellinger K, Tourtellotte WW. Human brain D1 and D2 dopamine receptors in schizophrenia, Alzheimer's, Parkinson's, and Huntington's diseases. Neuropsychopharmacology 1(1):5–15; 1987. (PubMed)

- 4. Stoof JC, Kebabian JW. Two dopamine receptors: biochemistry, physiology and pharmacology. Life Sci 35 (23):2281–2296; 1984. (PubMed)
- 5. George SR, Watanabe M, Di Paolo T, Falardeau P, Labrie F, Seeman P. The functional state of the dopamine receptor in the anterior pituitary is in the high affinity form. Endocrinology 117(2):690–697; 1985. (PubMed)
- Gehlert DR, Wamsley JK. Autoradiographic localization of [3H]sulpiride binding sites in the rat brain. Eur J Pharmacol 98(2):311–312; 1984. (PubMed)
- Lidow MS, Goldman-Rakic PS, Rakic P, Innis RB. Dopamine D2 receptors in the cerebral cortex: distribution and pharmacological characterization with [3H]raclopride. Proc Natl Acad Sci U S A 86(16):6412–6416; 1989. (PubMed)
- 8. Brucke T, Tsai YF, McLellan C, Singhanyom W, Kung HF, Cohen RM, Chiueh CC. In vitro binding properties and autoradiographic imaging of 3-iodobenzamide ([125I]-IBZM): a potential imaging ligand for D-2 dopamine receptors in SPECT. Life Sci 42(21):2097–2104; 1988. (PubMed)
- Kessler RM, Ansari MS, Schmidt DE, de Paulis T, Clanton JA, Innis R, al-Tikriti M, Manning RG, Gillespie D. High affinity dopamine D2 receptor radioligands.
   [125l]epidepride, a potent and specific radioligand for the characterization of striatal and extrastriatal dopamine D2 receptors. Life Sci 49(8):617–628; 1991. (PubMed)
- Mukherjee J, Yang ZY, Das MK, Brown T. Fluorinated benzamide neuroleptics--III. Development of (S)-N-[(1-allyl-2-pyrrolidinyl)methyl]-5-(3-[18F]fluoropropyl)-2, 3-dimethoxybenzamide as an improved dopamine D-2 receptor tracer. Nucl Med Biol 22(3):283–296; 1995. (PubMed)
- 11. Grunder G, Landvogt C, Vernaleken I, Buchholz HG, Ondracek J, Siessmeier T, Hartter S, Schreckenberger M, Stoeter P, Hiemke C, et al. 2005.
- 12. Mukherjee J, Christian BT, Narayanan TK, Shi B, Collins D. Measurement of d-amphetamine-induced effects on the binding of dopamine D-2/D-3 receptor radioligand, 18F-fallypride in extrastriatal brain regions in non-human primates using PET. Brain Res 1032(1-2):77–84; 2005. (PubMed)
- 13. 13. Riccardi, P., R. Li, M.S. Ansari, D. Zald, S. Park, B. Dawant, S. Anderson, M. Doop, N. Woodward, E. Schoenberg, D. Schmidt, R. Baldwin, and R. Kessler, Amphetamine-Induced Displacement of [(18)F] Fallypride in Striatum and Extrastriatal Regions in Humans. Neuropsychopharmacology, 2006;31:1016-1026.
- 14. Gardner B, Strange PG. Agonist action at D2(long) dopamine receptors: ligand binding and functional assays. Br J Pharmacol 124(5):978–984; 1998. (PubMed)
- 15. Lahti RA, Mutin A, Cochrane EV, Tepper PG, Dijkstra D, Wikstrom H, Tamminga CA. Affinities and intrinsic activities of dopamine receptor agonists for the hD21 and hD4.4 receptors. Eur J Pharmacol 301(1-3):R11–R13; 1996. (PubMed)
- Seeman P, Watanabe M, Grigoriadis D, Tedesco JL, George SR, Svensson U, Nilsson JL, Neumeyer JL. Dopamine D2 receptor binding sites for agonists. A tetrahedral model. Mol Pharmacol 28(5):391–399; 1985. (PubMed)
- 17. Neumeyer JL, Neustadt BR, Oh KH, Weinhardt KK, Boyce CB, Rosenberg FJ, Teiger DG. Aporphines. 8. Total synthesis and pharmacological evaluation of (plus or minus)-apomorphine, (plus or minus)-apocodeine, (plus or minus)-N-n-propylnorapomorphine, and (plus or minus)-N-n-propylnorapocodeine. J Med Chem 16(11):1223–1228; 1973. (PubMed)
- 18. Wong A, Hwang SM, McDevitt P, McNulty D, Stadel JM, Johanson K. Studies on alpha v beta 3/ligand interactions using a [3H]SK&F-107260 binding assay. Mol Pharmacol 50(3):529–537; 1996. (PubMed)
- 19. Sibley DR, De Lean A, Creese I. Anterior pituitary dopamine receptors. Demonstration of interconvertible high and low affinity states of the D-2 dopamine receptor. J Biol Chem 257(11):6351–6361; 1982. (PubMed)
- 20. Narendran R, Hwang DR, Slifstein M, Talbot PS, Erritzoe D, Huang Y, Cooper TB, Martinez D, Kegeles LS, Abi-Dargham A, et al. In vivo vulnerability to competition by endogenous dopamine: comparison of the D2 receptor agonist radiotracer (-)-N-[11C]propyl-norapomorphine ([11C]NPA) with the D2 receptor antagonist radiotracer [11C]-raclopride. Synapse 52(3):188–208; 2004. (PubMed)

21. Narendran R, Hwang DR, Slifstein M, Hwang Y, Huang Y, Ekelund J, Guillin O, Scher E, Martinez D, Laruelle M. Measurement of the proportion of D2 receptors configured in state of high affinity for agonists in vivo: a positron emission tomography study using [11C]N-propyl-norapomorphine and [11C]raclopride in baboons. J Pharmacol Exp Ther 315(1):80–90; 2005. (PubMed)